REVIEW ARTICLE

Taxonomy, distribution, biology and conservation of vulnerable snow trout *Schizothorax richardsonii* (Actinopterygii: Cyprinidae: Schizothoracinae) in the Himalayan and sub-Himalayan region: A review

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Abstract

Fish are a significant component of the aquatic environment and are crucial in supporting and maintaining the entire ecological system, thereby making freshwater systems more resilient and sustainable. The structure of a fish community is influenced by biotic interactions and abiotic variables. In recent years, spanning from the last 5 to 10 years, conservationists and commercial fishermen have reported a significant decline in freshwater fisheries. The decline can be attributed to several factors, including the introduction of exotic species, construction of dams, anthropogenic activities, illegal fishing, and overfishing. Schizothorax richardsonii, is a highly valued freshwater fish, inhabiting the foothills of the Himalayan regions. The declines of S. richardsonii in the Himalayan region are more than 90% in some areas and the predicted overall reduction is inferred to be around 50% with similar rates. Withstanding all the variations in biotic and abiotic factors, the species is expanding its range of altitude and evolving according to the climate-driven shifts in the Himalayas and apparently, its current distributional range would be lost over time. In Indo-Himalayan ranges, there is a lack of conservative measures and as a result, despite its ecological importance, snow trout has been given a vulnerable status by IUCN 2020. This review aims to provide descriptive biology and discuss the anthropogenic impact on the population structure of snow trout and its conservation.

Keywords: Aquaculture, IUCN, Morphometric, Protection.

INTRODUCTION

Among all vertebrates, fishes have the highest species diversity with about 36,617 valid species in 5,246 genera out of which 18,597 are found in freshwater ecosystems (Fricke 2023). The freshwater system is one of the major biomes on earth consisting of 68.7% of cold water fisheries inhabiting mainly the Himalayas. The cold water fisheries are established as potential candidates for aquaculture and providing economical support in the Himalayan uplands. The most abundant and important cold water fishes belong to the order Cyprinidontiformes with 23 families including Cyprinidae (Fricke et al. 2023). Among these, the subfamily Schizothoracinae could serve as a model species for assessment of thermal stress due to its ability to survive near-zero temperatures in streams of lesser and greater

Himalayas. They have evolved analogously to resemble trout, hence popularly known as snow trout (Kapila et al. 2002; Sharma et al. 2021a). Schizothorax richardsonii (Gray, 1832) is an endemic freshwater fish inhabiting rivers and streams of the Himalayan and sub-Himalayan regions in South Asia, Nepal, Bhutan, India, and Pakistan (Agarwal et al. 2009; Sarma et al. 2018; Agarwal 2019; Das et al. 2019; Fricke et al. 2023). Several taxonomical uncertainties lead to unreliable estimation of the location-specific distribution of this species however, the morphometric and molecular analyses based on complete mitogenomes analysis (Zhang et al. 2016) may serve as a probable taxonomic tool for the identification and classification of S. richardsonii (Shabanum & Dhanze 2016).

In the Himalayan riverine systems, the species

richness of the fish is influenced by physicochemical and variable factors like velocity, temperature, water discharge (Oberdoff et al. 1995), numerous anthropogenic threats, and climate change (Nag & Bhatacharjee 2002; Gupta et al. 2017). Intense human intervention has resulted in habitat loss and degradation of the freshwater ecosystem thus affecting the fish species, especially in regions with high water demand (Sarkar et al. 2012). Also, the introduction of exotic species intensifies the significant loss of the species with irreversible changes in its natural population (Dudgeon et al. 2006). As a consequence of unnatural deaths, S. richardsonii has significantly declined with other 4.27% of cold-water fish species (Cyprinion semiplotum, Botia rostrata, Aborichthys boutanensis, and Pseudecheneis sirenica) and falls under the vulnerable category of the IUCN red list (Sharma 2019; IUCN 2020; Dey et al. 2021). The reassessment of the conservation status of the S. richardsonii is critically important as it is getting coevolved with changing patterns of the Himalayan geomorphology (Sharma et al. 2021a). Thus, it leads to a continuous struggle for policymakers to design targeted and site-specific conservation plans for snow trout. Although, various protection amendments for freshwater biodiversity have been implemented at the global level (Linke et al. 2012; Heino et al. 2016). Here, much literature has been cited in order to dig out the ecology, biology, conservation, and population dynamics of Schizothorax richardsonii raising a huge concern for the conservation of this species before it becomes 'endangered'. There are several ongoing researches in the field of aquaculture explaining the morphology, biology, fishery, and culture of S. richardsonii, but in order promote sustenance and conservation, to consolidated review report needs to be amended. In this review, we have briefly highlighted the habitat, geographical distribution, feeding habits, migratory patterns, reproduction, commercial exploitation and also emphasized on its vulnerability (Fig. 1). We have also discussed briefly the life history of snow

trout that can be associated with population losses and vulnerability of the fish and several new threats that have emerged and are likely to jeopardize the future survival of this important fish species.

Taxonomy: There are 15 genera in the sub-family Schizothoracinae (family Cyprinidae) with a total of 100 species and 63 valid species under the genus Schizothorax (Zhang et al. 2018).

From Central Asia, 34 species have been reported, out of which 28 species of Schizothorax are found in the Indo-Himalayan region (*S*. richardsonii, S. curvifrons, S. esocinus, S. kumaonensis. S. hugelli, S. labiatus, S. longipinnis, S. micropogon, S. molesworthi, S. nasus, S. niger, S. planifrons, S. plagiostomus, S. sinuatus and S. progastus) (Misra 2003). On the basis of panoramic study of morphological datasets, currently Schizothoracinae is prorated into three divisions as 1) The primitive group (genera *Aspiorhynchus* and *Schizothorax*) 2) specialized group (genera Diptychus, Gymnodiptychus, and Ptychobarbus) and 3) highly specialized group (genera Chuanchia, Gymnocypris, Herzensteinia. Oxygymnocypris, Platypharodon, and Schizopygopsis) (He et al. 2004).

The classification of Schizothoracinae and the taxonomical status and evolutionary lineage of the genus Schizothorax is dubious (Jhingran 1991; Ahmed et al. 2014). This genus comprises of a collection of species that share a striking degree of morphological similarity and are frequently quite challenging to differentiate purely on exterior morphological characteristics (Chandra et al. 2012). The amalgamation of conventional taxonomical methods (morphological and osteological characters) and recent amelioration in DNA barcoding has paved the way for in-depth examination and identification of fisheries worldwide (Ward et al. 2005; Bhattacharya et al. 2016). Researchers have employed critical analyses of both Cyt *b*(Cytochrome b) and *COI*(Cytochrome oxidase subunit-1) sequences to clarify the molecular diversity of genus Schizothorax (Chandra et al. 2012; Akhtar et al. 2016; Shen et al. 2019; Ma et al. 2020).

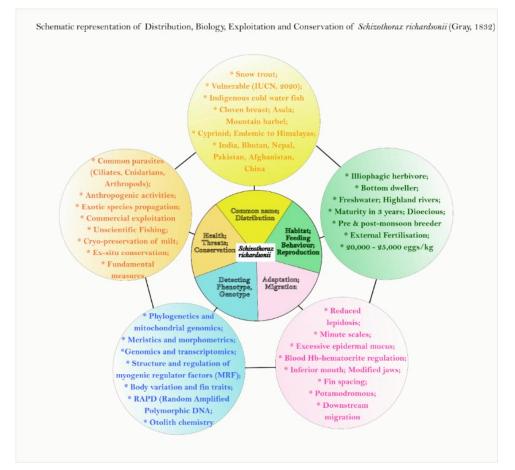


Fig.1. Brief summary of general biology, geographical distribution of Schizothorax richardsonii (Gray, 1832).

Chandra et al. (2012) found high genetic divergence across species and low genetic divergence within species, proving the effectiveness of COI barcodes molecular characterization of Schizothorax for species. 17 Schizothorax specimens from Neelum Jhelum Rivers, Pakistan was molecularly and identified based on COI (Akhtar et al. 2016). Recently, Ma et al. (2020) employed COI geneto successfully identify 185 specimens belonging to 11 Schizothorax species and Cyt. b gene to identify 264 specimens belonging to 23 Schizothorax species. Further, the taxonomic discrepancies, existence of cryptic lineages, and the emergence of new species in the genus Schizothorax can be resolved by examining genetic divergence between sequences and performing phylogenetic analysis (DeSalle et al. 2019).

Morphology: This fish species have a streamlined body covered with minute scales. The skin is smooth and soft to enable cutaneous respiration. Snowtrout possess blunt head with wide and levelled interorbital space and two pairs of barbels (Sharma & Mehta 2010). The pharyngeal teeth are arranged in three rows (4,3,2/2,3,4) and the lips are keratinised (Pandey & Nautival 1997). The inferior mouth is shorter than the upper jaw with a band of hard papillated adhesive organ. Adhesive organ is composed of keratinized epidermis cells (Das & Nag 2008) and aid in adhesion to rocky substrates. This keratinization of the peripheral row cells of adhesive organ might protect this structure against mechanical damage. The peripheral epidermal cells are made up of tonofilaments and mucous granules and the callus forms cavities encircled by tuberculated borders which support adhesion. The pectoral fins are shorter than the head and dorsal fin arises at pelvic fin anteriorly. The dorsal spine is serrated, strong and bony at the posterior end. On either side of the vent and anal fin, there is a sheath formed of tilted row of scales (Sharma & Mehta 2010).

Distribution in India and neighbouring countries: From the late Miocene epoch, a series of geological events have diversified and expanded the speciation of the genus Schizothorax Heckel, 1838 from its primitive ancestors and localized them in the great Himalayas with a very wide distribution. Since then, Schizothorax is the most dominant genus in the family Cyprinidae, inhabiting streams and rivers in the Himalayan and sub-Himalayan regions. Its distribution has been reported from India, Bhutan, Nepal, Pakistan, and Afghanistan. The Himalayan mountain range extends up to a 2400 km long arc from east-southeast to west-northwest (Wadia 1931). The westernmost ranges are distinctly bounded by the river Indus, the easternmost is demarcated by the Brahmaputra, and the north and south are the lower limits of Gangetic plains (Goswami et al. 2013, Sharma et al. 2021a). Schizothorax richardsonii distribution has been reported from Central, Eastern, and Western regions and also extends from Northwest Himalayas to Northeast Himalayas (Sehgal 1988; Shrestah 2002). Snow trout is regarded as a potential model organism to study climate-induced changes across the Himalayan riverscape as it has co-evolved with the changing patterns of the geomorphology (Sharma et al. 2021a). In a study, empirical findings revealed that snow trout would strongly persuade to mid elevated zones of the Himalayas due to climatedriven changes. Here, snow trout was found to be cooccurring with the genera viz., Tor, Neolissochilus, Barilius, and Nemacheilus (Majhi et al. 2013; Sharma et al. 2015). Recent reports predicted that in the near future, considerable range shifts and dramatic shrinkage in the population of snow trout could occur throughout the eastern Himalayas as compared to the western and central Himalayas reasonably due to the collapse of climatically controlled habitat (Sharma et al. 2021a).

Length-weightrelationships(LWRs):Theenvironmentinfluencesthelength-weightrelationshipinfishesandtheserelationshipsusefulindifferentiatingsmalltaxonomicunits

computing the shift from the expected weight for the length of the fish as an indication of well-being of the fish (Chonder 1972). Researchers attempt to unveil the length-weight relationship and relative condition factor (Kn) to reflect the growth pattern of fish under study (Jhingran 1952). In the cases where the fish retains the same shape with constant specific gravity, it shows isometric (value of exponent 'b'= 3.0). A value less than 3.0 shows that fish becomes lighter and greater than 3.0 indicates heavier for a particular length as it increases in size (Wooton 1990). Particularly, the analysis of the length-weight data for Schizothorax richardsonii reveals that the value of 'b' is close to 3 and fish shows the isometric growth (Mohan 2006, Sharma & Dhanze 2010). In some recent studies, this fish showed negative allometric growth indicating a changing environment (Mehmood et al., 2021; Malik et al. 2021; Khanal et al. 2023). The intra-specific variation in the value of 'b' or 'n' is under the influence of several factors such as seasonal, physiological condition, sex, gonadal development, and nutritive condition of the environment. Furthermore, the 'Kn' value is a physiological indicator of the general well-being of any fish living in a given environment (Kumari et al. 2006) where 'K' value > 1 indicates that the general well-being of the fish is good whereas its values < 1in some of the age groups indicate some issues in their habitat. In case of snowtrout, 'Kn' values have been reported almost constant and close to 1 in all size groups (Kumari et al. 2006; Sharma & Dhanze 2010; Neupane 2021).

Food and feeding habits: There are numerous studies indicating that the food habits of this species vary with the size and life cycle stages (Sehgal & Sar 1989; Sehgal 1990; Malik et al. 2018). In one of the tributaries of Ganga, and a few streams of Kashmir, the fish is a periphytic feeder, feeding on Bacillariophyceae (highly), Chlorophyceae, Cyanophyceae, detritus, and sand in this very preferential order and its diet is reported to have 75% plant matter and 25% of animal matter (Dipteran larvae, mayfly nymph, caddisflies larvae) (Shekhar et al. 2013; Koundal et al. 2016). Bacillariophyceae forms a major percentage of food throughout the year but decreases quantitatively during May-August whereas Chlorophyceae being the second major component of fish food increases quantitatively during May-August which may be due to the rise in water temperature and algal production (Shekhar et al. 2013). Apparently, feeding upon plants, aquatic insects, larvae, and nymphs results in alterations in the length of their gut during different stages of the life cycle i.e. longer in adults and shorter in juveniles (Koundal et al. 2013; Malik et al. 2018). The ventral position of the mouth is correlated with the bottomfeeding habit of the fish. With the help of a rasping mouth, it feeds on microbiota present on rocks and stones and on green and blue green algae.

In S. richardsonii, quantitative feeding in both sexes is low during monsoon and spring whereas to inflate the heat it is high in summer (May-June) due to high metabolism, rise in algal production, and allogenic inflow (Shekhar et al. 2013; Malik et al. 2018). Likewise, a slight increase in food intake during February-March and low quantitative feeding during April-May could be due to breeding (Jan et al. 1970; Jyoti & Malhotra 1975; Madhwal et al. 1984; Sunder 1985). After winter, the stimuli during the maturation of gonads and post-spawning activity increase the efficiency of the fish to build more energy reserves. A generalized picture that emerges from various studies indicates that values of the Enterosomatic index (ESI) and Gastrosomatic index (GSI) of this fish species are low while spawning and their index values increase during preand post-spawning periods i.e. June and October, respectively (Shekhar et al. 2013; Koundal et al. 2013; Malik et al. 2018).

Reproductive biology (Sexual dimorphism, Sex ratio, gonad maturity stages, spawning season, fecundity): Sexual dimorphism is quite evident where mature males are brighter in colour with straight ventral profile whereas mature females are with distended bellies (Ciji et al. 2021). During the initial growth phase, the ovaries are paired, flaccid, and translucent, but lately, in summers they become distended and fully matured with ripened ova (2.5-4.3mm) (Joshi 2004; Gandotra et al. 2009; Gandotra 2009). The quality of gametes and the activity of gonads are highly variable depending on multiple external factors such as feeding patterns and ambient temperature (Sabha et al. 2011; Bahuguna et al. 2011). The success rate of fertilization mainly depends on the qualitative and quantitative characteristics of semen (Bobe & Labbe 2010). The fish breed twice annually one with the onset of summer (April-May) in upstream water with a sudden rise in temperature (15-19°C) and the other during the monsoon (Sehgal 1988, Raizada et al. 1983).

Schizothorax richardsonii is characterized by slow growth and late sexual maturity in their surrounding rigorous environment (Chen & Cao 2000; Mohan 2005). Eggs are large-sized, laid in shallow pools, and remain attached to the substratum until the fry hatches. There is a linear relationship between the number of eggs and an increase in body lengthweight or ovary's length-weight (Table 1) (Jan et al. 2014). In S. richardsonii, low fecundity was recorded within a range from 20,000-25,000 eggs/body weight (kg) (Shahnawaz & Pandey 2016). A long-term study was carried out from 1996-98 and 2016-19 that covered the fecundity rates of the Schizothorax during the timeline (construction to commissioning) of a hydroelectric power project at Uttarakhand, India. The study revealed a constant rate of fecundity and survival attributes due to its unique adaptive skills and shifting of the breeding grounds (Thapliyal et al. 2019).

Clear shallow water, gravelly sandy beds, with very feeble water flow are characteristics of spawning grounds of this species (Ali & Pandey 2016). There are studies exhibiting variation in the spawning cycles of *S. richardsonii*, describing its extemporary behaviour with respect to the water temperature, food availability, and photoperiodism (Bahuguna et al. 2011). Past studies showed that species spawn once or twice (April-May, July-August) or multiple times

Species name	Geographical Occurrence (Himalayas)	(GSI) Highest- Lowest	Fecundity (eggs /g body weight).	References
Schizothorax richardsonii Growth (cm/year): 10.0-13.0 Size (cm): 13.0-16.0 Maturity age(year): 2- 3 years	Throughout the Himalayas	-	25.0-80.0	Basavaraja (2007); Bhatt & Pandit (2016)
	Kashmir	-	2600-16605	Das & Koul (1965)
	Kashmir	-	6720-27500	Rampal (1967)
	Sindh Nallah, Kashmir	-	2598-27312	Mir (1979)
	Alkananda from Garhwal Himalayas	Sep-April	3832 -5310	Misra (1982); Joshi et al. (2016)
	Sindh and Telbal Nallahs	-	2598-27846	Qadri et al. (1983)
	Garhwal region	Jan-Oct	1578-14316	Badola & Singh (1984); Baloni & Tilak (1985); Thapliyalal et al. (2012)
	Neeru Nallah, Bhadarwah	-	970-6035	Shekhar (1990)
	Kumaon river	-	8216-48490	Singh (1990)
	Kumaon river	-	1311-12702	Bhatt & Pathak (1992)
	River Gaula, district Kumaon,	July-Oct, Jan-Feb	2248-8726	Mohan (2005); Roy (2011)
	Rajouri district, J&K		3032-13321	Gandotra (2009)
	Nallu River, Lalitpur District	Sept – Nov, May	6294 to 67083	Wagle (2015)

Table 1. Intrinsic characteristics of Schizothorax richardsoni.

depending upon different environmental conditions and elevations of the rivers/streams (Bishat & Joshi 1975; Jhingran & Sehgal 1978). The spawning season is determined on the basis of gonadosomatic indices (GSI), ova diameter, occurrence of brooders, eggs, and larval stages in the spawning grounds. In elite cases, the species have the ability to regulate their spawning season through migratory patterns due to dramatic changes in the thermal regime of each drainage and flood conditions created in mountain streams due to snow melting (Singh 1990; Bhatt & Pathak 1992; Rai et al. 2002; Joshi 2004). The antagonistic relationship between spawning and gonadotrophic hormones reported earlier stated that there is a decline in the levels of spermatocrit and sperm density with the onset of spawning in these fishes (Agarwal & Raghuvanshi 2009; Ciji et al. 2021).

Migratory patterns: The snow trout has potamodromous migratory behaviour and its exact migratory distance is yet to be ascertained. In rivers like Beas and Sutlej, *S. richardsonii* undertakes long-

distance upstream migration from the Greater Himalayas to the Sivalik regions as it prefers underground water for spawning (Rai et al. 2002). However, in the upper Beas, the species spawns in monsoon when the water temperature is warm enough and continuously migrates downstream to spawn during winters (Petr et al. 2002). When the temperature increases in the summer and dissolved oxygen decreases, snow trout cannot be collected at lower elevations while in the winter season, it has been recorded from the same stream sites. So, the fish spend most of their life span in the upland waters but migrate downstream for breeding, feeding, and spawning during over-wintering, intense human intervention, and natural disasters (Jackson & Marmulla 2001).

Acclimation regimes: This species is specialized for the torrential streams and has co-evolved with the changing patterns of Himalayan geomorphology. The thermal range reported for Indian snow trout ranges from 0-27 °C (Sharma & Mehta 2010) and the fluctuations in ambient temperature are responsible

for influencing the spawning, ovulation, body morphology, and hematology (Qadri et al. 1983; Kapila et al. 2002). Due to remarkable uniformity in the body contours, S. richardsonii thrives well in the shallow zone of the fast water currents exhibiting greater variability corresponding to environmental fluctuations (Negi & Negi 2010; Shekhar et al. 2013; Wagle 2014). Lack of lipidosis on the ventral side of the body, reduced scales, and horizontally placed paired fins provide adhesion against the substratum for feeding (Das & Nag 2008; Sharma 2010; Hayden et al. 2014; Singh & Bisht 2017). The feeding structures of S. richardsonii including epithelia of the lips, rostral cap, adhesive pad, and lower lip undergo modifications to form keratinized, hard and sharp edges on the labial plate that confers the adaptation for its herbivorous mode of feeding (Singh et al. 2014; Koundal et al. 2016).

Habitat monitoring experiments suggested that the species could survive from hypoxia in winters, as the lowering in blood haemoglobin-haematocrit levels could limit the oxygen carrying capacity, with probable reciprocations in aerobic metabolism and thermal tolerance (Kamalam et al. 2019). The tolerance levels of fish undergoes irreversible and reversible changes with life stages, seasons, acclimation regimes, and nutrition conditions (Beitinger & Bennett 2000; Wagle 2015; Joshi et al. 2017). Schizothorax has survived through the evolutionary age of the Himalayas due to its adaptive skills and has acquired characters making it suitable for living in the fast-flowing and torrential hill streams (Rajput et al. 2013). Thus, snow trout has a great adaptive capacity to grow and reproduce in fast-flowing torrential river systems (Wagle 2015).

Phenotypic and genotypic provisions: By modification in the physiology and behaviour of snow trout, adaptive response to environmental change is allowed by the phenotypic plasticity of this fish which leads to changes in their morphological features, reproduction, or survival that moderate the particular effects of environmental factors like temperature, water current velocity, and latitude

(Meyer 1987). For stock identification, many tools like meristic, morphometric, parasites as natural tags, traditional tags, otolith chemistry, molecular genetics, and electronic tags have been used (Negi et al. 2015; Regmi 2019). The different environmental features of habitats including water depth, temperature, velocity, and availability of different substrates affect the morphological features of inhabitant fishes (Rajput et al. 2013). Fin size and body shape specifically are two imperative morphological characters that contrast greatly in lentic vs. lotic habitats (Douglas & Matthews 1992). Due to its presence across a broad range of physicochemical stream conditions, with a wide range of environmental and physiological tolerances, snow trout is a habitat generalist (Rajput et al. 2013).

The mitochondrial genome of S. richardsonii is 16,592 bp in length and comprises 22 tRNAs, 2 rRNA genes, 13 protein coding genes, and one putative control region, which consists of a microsatellite; (TA)13 that exists between 16.469-16,494 bp (Goel et al. 2014). Recently, Next Generation Sequencing technology has become a tool to study the genetic behaviour of the fish species in response to their environment (Talwar et al. 2018). With the help of RNA sequencing of the liver tissue of the snow trout, a reference transcriptome database has been generated using Illumina HiSeq 2000 platform. Annotated blast matches revealed that differentially expressed transcripts correspond to chaperones and molecular critical pathways, previously shown to be important for thermal stress in other fish species (Barat et al. 2016). The transcripts identified as solute carrier transporter genes are categorized under 13 different protein families that play an integral role in cellular acclimation response (Barat et al. 2019). Slow growth in this important Himalayan cyprinid can be deciphered by the molecular characterization of the structure and regulation of the myogenic regulatory factors (MRFs; myod, myf5, myogenin (myog), myf6/herculin/mrf4) as through the processes of cell determination and differentiation, MRFs play an important role in muscle growth (Rajesh et al. 2019).

The complementary study of phenotypic and genotypic variations can serve as an attempt at stock assessment of snow trout for rational exploitation, conducting conservation, and sustainable management measures. In a study to understand the environmental impact of the morphology of S. richardsonii, strong correlations were suggested between environmental and morphological features (Rajput et al. 2013). The different body and fin traits in snow trout are shown to be adapted for a specific habitat and also a negative correlation between body size and fin morphology has been suggested (Rajput et al. 2013). The relationship between AFL and A-CFBL (anal to caudal fin base length) in the species under investigation was found to be a significant feature as relatively smaller AFL and longer A-CFBL in S. richardsonii serve to distinguish it from S. plagiostomus (Pandey & Nautiyal 1997).

detailed А more analysis of multiple morphometric characters showed that characters such as standard length and pre-anal distance correlate with the total body length while dorsal fin and depth of anal fin correlated least with the total body length in a study conducted on samples collected from River Yamuna of the Uttarkashi district of Uttarakhand, India (Negi & Negi, 2010). They also showed that characters such as standard length, predorsal distance, preventral distance, preanal distance, maximum body depth, minimum body depth, caudal peduncle length, head length (HL), head width, pelvic and anal fin, pectoral, pelvic (ventral) increase in direct proportion with each other. On the other hand, postorbital distance and head depth are shown to highly correlate with the HL while eye diameter correlates least with the HL in snow trout (Negi & Negi, 2010). These morphometric characters are categorized into genetically often controlled (minimum variability), environmentally controlled (maximum variability), and intermediate characters (slightly controlled by environment). A number of morphometric characters that included HL, pre dorsal distance (PrDD), post dorsal distance (PoDD), depth

of anal fin (DAF), length of anal fin (LAF), depth of dorsal fin (DDF), length of dorsal fin (LDF), length of pelvic fin (LPF), length of caudal fin (LCF), distance between pectoral and pelvic fin (DPP), maximum body depth (MBD), Minimum body depth (MiBD), head depth, eye diameter, pre-orbital and post orbital distance stated that 90% variation in morphometric characters of this species populations from Uttarakashi, India are genetically controlled opposing the fact that the maximum no. of morphometry are environment-biased (Negi & Negi 2010). Even the intraspecific variation investigation of the S. richardsonii, established on the basis of morphometric characters from rivers in the Western and Central Indian Himalayas displayed significant phenotypic heterogeneity which could be due to the local ecological conditions (Mir et al. 2013). Furthermore, the variation in the morphometric characteristics was also measured by using the Wilks λ tests of discriminant analysis showed that there are noteworthy variations in morphometric characteristics among few inhabitants due to phenotypic plasticity in response to uncommon hydrological conditions like variation in alkalinity, temperature and closeness may be due to similar habitat attributes (Mir et al. 2013).

Using RAPD (random amplified polymorphic DNA technique), the genetic heterogeneity was found to be accelerated with respect to isolation and distance in sampling sites (Kapila et al. 2006). In a recent study, independent and dependent variables were environmental analysed using correlation coefficient and descriptive statistical parameters where the total length of fish species was described high. Lohani et al. (2020)the environmentally controlled variability in the specimens. So, the phenotypic and genotypic characters can provide insights in the phenotypic plasticity and conserved patterns of the fish species at the surroundings need to be studied throughout the time.

Concurrent pathogenesis: Very less information about interactions between *S. richardsonii* and

Species	Host	Site of infection	Locality
<i>Myxobolus himalayaensis</i> (Ahmed et al. 2019)	Schizothorax richardsonii	Gill filament	River Poonch (Madiana)
<i>M. kashmirensis</i> (Dar et al., 2017 b)	Schizothorax esocinus	Gills	J&K (India)
<i>M. chushii</i> (Dar et al. 2017 a)	Schizothorax niger	Gill lamellae	J&K (India)
<i>M. nigerae</i> (Dar et al. 2016)	Schizothorax niger	Gill lamellae	J&K (India)
<i>M. linzhiensis</i> (Li et al. 2017)	Schizothorax oconnori	Gill	China
Argulus spp. (Mallik et al. 2010)	Schizothorax richardsonii	Caudal and anal fins	Bhimtal (India)
Ichthyophthirius multifiliis (Mallik et al. 2015)	Schizothorax richardsonii	Dorsal body surface, caudal fins	Champawat, Uttrakhand, (India)

Table 2. List of ectoparasites and endoparasites known for their infection in various species of Schizothorax.

pathogens has been recorded, as a consequence, the concern for fish health remains subtle. There is a higher risk of diseases in cold water fish due to significant temperature fluctuations, which in the long run affects their population and diversity. A wide range of ciliates, cnidarians, and arthropods act as parasites that are responsible for a high infection rate and may increase the mortality of this fish species (Table 2) (Mallik et al. 2010; Ahmed et al. 2019). The majority of the studies have indicated that the known arthropod Argulus sp. commonly called as fish louse have become a major threat for the health management and aquatic crop production for fish in tropical and temperate regions. This ectoparasite mainly infested inland capture and culture fisheries of cyprinids, as temperatures of these water bodies provide an optimum environment for its life-cycle (Mallik et al. 2010). In Argulus infestation (Aurgulosis), the mortality due to re-infection increases, reportedly in 2008, stocks found to be infested due to secondary infection of this ectoparasite which became a major cause for low productivity of S. richardsonii and T. putitora in Uttrakhand, India. The infected S. richardsonii was recorded 51.2 %, whereas abundance and mean intensity of infestations were 1.05 and 2.06 respectively and the maximum prevalence (70.1%)was observed in September with the least in

December (Mallik et al. 2010). Similarly, the endoparasite *Myxobolus* sp. causes necrosis. hyperplasia, and hypertrophy in the plasmodia of type FV2 of gill filament in its host fish species snow trout (Ahmed et al. 2019). Myxobolus sp. infection was examined in *Schizothorax richardsonii* with a prevalence of 26.25%. Further, investigation of a temperature-dependent ciliate parasite. Ichthyophthirius multifiliis that causes pinhead sized white spots (0.4-0.8mm) on the dorsal body surface and caudal fins of this fish species was reported (Mallik et al. 2015). Temperature-dependent infection pattern was noted with the maximum prevalence of 84.8% in month of July. Recently, a parasite Saprolegnia parasitica was isolated and identified from infected eggs and adults that was mutually present with S. australis in early stages of snow trout (Jen 2008). Another pathogenic bacterial sp., Aeromonas hydrophila is known to infect the hematopoietic tissue in spleen and kidney in this fish species (Uzma et al. 2020).

Fish species are known to harbour anti-microbial peptides (AMP) within the gut, as these AMP-like molecules enable the host's innate immune system to suppress various infectious microbes mainly bacteria and viruses. In 2014, hepcidin-like anti-microbial peptide was amplified, cloned and characterized exclusively from *Schizothorax richardsonii* which

was found to be different from other cyprinids with a unique amino acid (Gln³⁰). The AMP molecule reported for its significant role in functioning of permeability of membrane molecules and thus enabling microbial retention (Chaturvedi et al. 2014). Threats: Among all the taxa, endemic freshwater fish species are the most vulnerable throughout the world due to intense human intervention in freshwater riverine system (Magurran 2009, Sarkar et al. 2012). Habitat alteration by dams, unscientific methods of fishing, illegal boulder/sand mining, poaching, indiscriminate fishing and introduction of alien species are largely responsible for the vulnerability of S. richardsonii (Singh & Bisht 2017). From past few decades, the human population in Indian Himalayas has been increasing at faster rates, and the energy requirement and electricity building projects are being forced into the Himalayan landscape to meet the demands. The government of India has issued policies to exploit the riverine system of the Indian Himalayas, which is hypothetically proven to cause serious damage to biodiversity and changes in the ecosystem (Pandit & Grumbiene 2012). The causes of vulnerability have been poorly understood and have not been regulated yet and thus the effects of threats cannot be addressed (Richter et al. 1997). An array of various factors such as habitat loss, habitat fragmentation, and overenthusiastic collection have increased the risk of extinction for this fish and many other fish species (Dudgeon et al. 2006). Certain favourable parameters of Himalayan rivers such as the perennial flow of streams, steep gradients and stable rock banks are projecting them for hydroelectricity development thus posing the ecological assets and aquatic resources including fisheries under severe threat (Joshi et al. 2017). Here, the extensive construction of dams and inappropriate regulation of the riverine system in the Himalayas, caused a decline in the species richness and altered the rates of fish diversity by hampering fish migration (Agarwal et al. 2009; Pandit & Grumbine 2012; Mali & Chutia 2017; Bhatt et al. 2017; Singh 2018; Agarwal et al. 2018). In a few cases, the hydro-power built over

Chenab rivers in the central Himalayas, has demolished their natural habitat by blocking the migratory routes of the snow trouts sp. (viz. *Schizothorax* richardsonii, Schizothorax plagiostomus, Schizothorax curvifrons) (Agarwal 1996, Agarwal 2001). Also, the increased siltation at bottoms in Pong Dam reservoir has affected the downstream fishing and also the catch percentage has been diminished specifically for Schizothorax richardsonii (Gray) (Sharma 2018). In Nepal, the Indrasarobar reservoir was formed by damming the Kulekhani River as part of long-term hydroelectric power dvelopment which led to diversion in the fish species composition and rapid decline in snow trout (Swar 1990). In spite of being a prime rheophilic species, S. richardsonii (Sharma et al. 2021b), has still not been considered while designing the dams in basins like Parvati, where flow fluctuations act as chief stressors for the native snow trout (Johal et al. 2021).

The unscientific methods used for fishing i.e., bleaching powder, insecticides, dynamiting, hammering and electric current are responsible for the mass mortality of all sizes of fish (Badoni et al. 2018). The introduction of alien species is one of the major threats to native fish populations. Common carp, introduced in the Kashmir has almost exterminated the indigenous schizothoracids of the Valley (Agarwal 2009). Likewise, the acceptance of brown trout as a sport fish in the Himalayan region poses superfluous reproductive pressure on the snow trout that warrants quantification through future research (Sharma et al. 2021b). Also, a rapid deterioration in the catches of this species in the Himalayas as the result of predatory brown trout preying upon their younger stages (Raina and Petr 1999; Petr et al. 2002). Due to climatic change, S. richardsonii has been predicted to lose its habitat by 16% in the coming 30 years and around 26% by 2070. Moreover, the declines of snow trout in the Himalayan region are more than 90% in some areas and the overall reduction is inferred to be <50% with similar rates predicted in the future (Sharma et al.

2021a).

The extraction of boulders, cobbles, gravels, and sand from the river bed alters the stream morphology and is further responsible for the disturbance of the breeding ground of fish species (Hassan et al. 2017). Degradation of habitat, inhibition of spawning migration, and intensive fishing pressure population have led to the rapid decrease of snow trout (Bruton 1995). Reports indicating the dramatic decline in growth and population size of snow trout in Ladakh (Sivakumar 2008) and Kashmir in the last five years was due to very slow growth, low fecundity, and early maturity of the species (Mir et al. 2012). Slow growth to maturity and short growth period and are the main constrictions deterring its growth and population (Mir et al. 2012). The International Union for the Conservation of Nature and Natural Resources (IUCN) developed the major classification system internationally used for the assessment of the threat status to each species and snow trout has been given the vulnerable status (Vishwanath 2018; IUCN 2020). Schizothorax richardsonii is considered as vulnerable according to IUCN 2020 and needs urgent fortification and proactive conservation efforts prior to its complete disappearance in most of its range.

Conservation: Conservation involves social, political, and economic imperatives, modification, and maintenance of the genetic identity and integrity of the species in their natural habitat with genetically sustainable fishery (Margules & Pressey 2000; Lakra et al. 2007). The conservative measure entails the identification and protection of important river stretches in the Himalayas that constitute crucial habitats of snow trout for conservation and restoration. measures need to follow These established by state environmental regulations authorities. ecologists, local communities, and conservationists. Instead, lack of management leads to various constraints such as the construction of dams, and commercial exploitation that were followed up irrespective of these integrated approaches. Till records, over 292 dams are commissioned proposed Himalayan or in

rivers which is a prominent habitat of snow trout (Pandit & Grumbine 2012). The dam constructions have led the distribution of snow trout to a very restricted area occupancy, moreover directed them to shift their habitat to mid-level altitudes. In the aquatic realms, snow trout is subjected to several biogeographical constraints such as physiographical barriers as well as by dendritic arrangement of riverine ecosystems (Rodriguez 2002; Olden et al. 2010; Singh 2019). The limited adaptability to human-intervened habitats and food resources could be the critical assessed factors that can hamper the population of snow trout in the Himalayas. Practically, the generation of vulnerability of this species could be avoided by conserving the habitats of native species before they become extinct (Moyle 1995). The availability of resources for conservation is very limited and needs to be rationally allocated (Sarkar & Bain 2007). Actions needed for the conservation of snow trout require site/area protection, resource, and habitat protection, habitat management, invasive species control, species recovery, reintroduction. species ex situ conservation, formal education, awareness, communication, and legal policies. Earlier, few policies have been implemented such as Ecosystembased fishery management (EBFM) supporting fisheries by addressing some of the unintentional consequences of fishing such as destruction of habitat, mortality of non-target species, and variations in the structure and function of ecosystems (Pikitch et al. 2004). In high-altitude regions of the Himalayas and Peninsula, the taxonomic study of all commercial fish species such as mahseer, snow trout, and common carp will be essential prerequisite steps to any broad program of resource conservation with a full checklist indicating the status of each species (Mijkherjee et al. 2002; Talwar et al. 2020). The random fishing in the natural environment could compromise the structure and function of the ecosystem if achieved without disturbing biodiversity. In order to balance both quantitative and selective fishing, the productive capacity of resources

for the snow trout must be increased (Zhou et al. 2010). In 2012, the Wildlife Institute of India suggested terminating a number of hydropower projects/dam construction to prevail in biodiversity conservation and management in the Himalayas. Very few commissioned projects were accepted with strict guidelines and regulations mainly including the reduced water flow speed (Rajvanshi et al. 2012). Research regarding population size, distribution and trends, life history and ecology, harvest, use, and livelihoods is needed for the conservation of snow trout. The river reaches, terrestrial protected areas that are managed by local stakeholders can act as management tools for biodiversity conservation as they have the potential to safeguard fish species (Gupta et al. 2015). During these studies, it has been found that encouraging vegetation throughout the streams has a greater impact on water alkalinity that is compatible with fish growth and survival. Likewise, some patches can be declared as sanctuaries by the Fishery Department to protect the fauna and the fish must be covered under the Schedule list of the Wildlife (Protection) Act (1972) (Sarkar et al. 2008; Raut et al. 2019). The biotechnology tools may establish completion of future demands that succeed with cryopreservation of brooders milt and introducing native population into the hatchery for artificial breeding (Agarwal et al. 2009; Rathore 2016; Hagedorn et al. 2018; Singh 2019) and stock protection from being totally eliminated due to natural disaster, sudden outbreak of disease, over exploitation, etc. (Agarwal 2011). Also, the technology of artificial fecundation of pondraised brooders and rearing of young ones in controlled conditions has been developed at research institutes (Singh 2016). In Indian cold-water fishes, induced spawning is difficult but artificial fertilization can be achieved by stripping mature brooders. A detailed study and extensive research must be performed before introducing any exotic species as it is reported that these species could challenge the lifestyle of native species and affect their abundance, in reality, the snow trout could be

pinned down in competitive limits of the natural habitat (Raina & Petr 1999). To achieve protection goals and for the conservation of aquatic ecosystems, the main challenge is the creation of markets wherein the value of fish killed is lower than living fish for consumption or sale where this outstrips sustainable limits (Everard and Kataria 2011). To restore this fishery, the first initiative for artificial was attempted propagation in Kashmir, and success was achieved in obtaining pure and healthy seeds of different species such as Schizothoraichthys niger, S. esocinus, S. curvifrons, S. micropogon and Schizothorax richardsonii through artificial fecundation (Joshi 2001; Sarma et al. 2018).

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REFERENCES

- Agarwal, N.K. 2001. Reproductive strategies of snow trout (*Schizothorax* sp.) with reference to environment of river Bhagirathi and Bhilangana of Tehri Garhwal. A final project report to Indian Council of Agriculture Research 78.
- Agarwal, N.K. 2008. Fish reproduction. APH Publishing Corporation, New Delhi. https://books.google.co.in/books?id=b0MryzLUQV0C
- Agarwal, N.K. 2011. Cryopreservation of fish semen. In: Himalayan Aquatic Biodiversity Conservation and New Tools in Biotechnology. Transmedia Publication, Uttarakhand 104-27.
- Agarwal, N.K.; Raghuvanshi, S.K. & Saini, V. 2009. Cryopreservation of snowtrout (*S. richardsonii*) milt as a means for propagation and ex-situ conservation of species. Fisheries Research 273-84.
- Agarwal, N.K.; Rawat, U.S. & Singh, G. 2019. Fish assemblages and habitat ecology of River Pinder in central Himalaya, India. Iranian Journal of Fisheries Sciences 18: 1-4.
- Agarwal, N.K.; Singh, G.; Singh, H.; Kumar, N. & Rawat, U. 2018. Ecological impacts of dams on the fish diversity of Bhagirathi River in Central Himalaya (India). Coldwater fisheries society of India 1: 74-84.

- Ahmad, S.M.; Bhat, F.A.; Balkhi, M.U.H. & Bhat, B.A. 2014. Mitochondrial DNA variability to explore the relationship complexity of Schizothoracine (Teleostei: Cyprinidae). Genetica 142(6): 507-516.
- Ahmad, M.; Malik, K.; Tariq, A.; Zhang, G.; Yaseen, G.; Rashid, N.; Sultana, S.; Zafar, M.; Ullah, K. & Khan, M.P.Z. 2018. Botany, ethnomedicines, phytochemistry and pharmacology of Himalayan paeony (*Paeonia emodi Royle*). Journal of Ethnopharmacology 220: 197-219.
- Ahmed, I.; Ahmad, I.; Dar, S.A.; Awas, M.; Kaur, H.; Ganai, B.A. & Shah, B.A. 2019. *Myxobolus himalayaensis* sp. nov. (Cnidaria: Myxozoa) parasiting *Schizothorax richardsonii* (Cyprinidae: Schizothoracinae) from River Poonch in North West Himalaya, India. Aquaculture Reports 14: 100192.
- Akhtar, T. & Ali, G. 2016. DNA barcoding of Schizothorax species from the Neelum and Jhelum Rivers of Azad Jammu and Kashmir. Mitochondrial DNA Part B 1(1): 934-936.
- Ali, S. & Pandey, N.N. 2016. Snowtrout in the Indian Himalaya ENVIS Newsletter on Himalayan Ecology 13.
- Badola, S.P. & Singh, H.R. 1984. Spawning of some important coldwater fish of the Garhwal Himalaya. Journal of the Bombay Natural History Society 811: 54-8.
- Badoni, A.K. 2018. Fish diversity of a small hill stream Dhundeshwer Gad of Uttarakhand (India) and need of its conservation. International Research Journal of Biological Sciences 7: 16-19.
- Bahuguna, S.N.; Tripathi, A.K. & Amir, K. 2011. Morphological and biochemical studies of the milt (Spermatozoa) of the snow-trout fish *Schizothorax richardsonii* (Gray). Environment Conservation Journal 12: 23-8.
- Baloni, S.P. & Tilak, R. 1985. Ecological observations on Schizothorax richardsonii (Gray). Journal of the Bombay Natural History Society 82: 581-5.
- Barat, A.; Sahoo, P.K.; Kumar, R.; Goel, C. & Singh, A.K. 2016. Transcriptional response to heat shock in liver of snow trout (*Schizothorax richardsonii*)-a vulnerable Himalayan Cyprinid fish. *Indian Journal of Fisheries* 16: 203-13.
- Barat, A.; Sahoo, P.K.; Kumar, R.; Goel, C.; Siva, C. & Ali, S. 2019. Data on solute carrier transporter genes of a threatened Himalayan fish species–*Schizothorax richardsonii*. Data in brief 23.
- Basavaraja, N. 2007. Freshwater fish seed resources in India. Assessment of freshwater fish seed resources for sustainable aquaculture 24: 267.

Beavan, R. 1877. Handbook of the freshwater fishes of

India. Reeve, London

- Beitinger, T.L. & Bennett, W.A. 2000. Quantification of the role of acclimation temperature in temperature tolerance of fishes. Environmental Biology of Fishes 58: 277-88.
- Bhatt, J.P. & Pandit, M.K. 2016. Endangered Golden mahseer *Tor putitora* Hamilton: a review of natural history. Reviews in Fish Biology and Fisheries 26: 25-38.
- Bhatt, J.P.; Tiwari, S. & Pandit, M.K. 2017. Environmental impact assessment of river valley projects in upper Teesta basin of Eastern Himalaya with special reference to fish conservation: a review. Impact Assessment and Project Appraisal 35: 340-50.
- Bhatt, S.D. & Pathak, J.K. 1992. Himalayan environment: water quality of the drainage basins. Shree Almora Book Depot, Uttarakhand
- Bhattacharya, M.; Sharma, A.R.; Patra, B.C.; Sharma, G.; Seo, E.M.; Nam, J.S.; Chakraborty, C. & Lee, S. S. 2016. DNA barcoding to fishes: current status and future directions. Mitochondrial DNA Part A 27(4): 2744-2752.
- Bisht, I. 1999. Adaptive modifications in the epidermis of the certain hill-stream fishes: a cytomorphological and histochemical investigation. (Doctoral dissertation, Ph.D. Thesis, Kumaun University Nainital, India).
- Bisht, J.S. & Joshi, M.L. 1975. Seasonal histological changes in the ovaries of a mountain stream teleost, *Schizothorax richardsonii* (Gray and Hard). Cells Tissues Organs 93: 512-525.
- Bobe, J. & Labbé, C. 2010. Egg and sperm quality in fish. General and comparative endocrinology 165(3): 535-548.
- Bruton, M.N. 1995. Have fishes had their chips? The dilemma of threatened fishes. Environmental biology of Fishes 43(1): 1-27.
- Chaturvedi, P.; Dhanik, M. & Pande, A. 2014. Characterization and structural analysis of hepcidin like antimicrobial peptide from *Schizothorax richardsonii* (Gray). The Protein Journal 33(1): 1-10.
- Chandra, S.; Barat, A.; Singh, M.; Singh, B. K. & Matura, R. 2012. DNA bar-coding of Indian coldwater fishes of genus *Schizothorax* (family: Cyprinidae) from Western Himalaya. World Journal of Fish and Marine Sciences 4(4): 430-435.
- Chen, Y.F. 2000. Schizothoracinae. Fauna Sinica, Osteichthyes, Cypriniformes: 273-388.
- Chonder, S.L. 1972. Length-weight relationship of mature female *Labeo gonius* (Hamilton-Buchanan) from the Keethan reservoir. Journal of Inland Fisheries Society of India 4: 216-217.
- Ciji, A.; Sharma, P.; Rajesh, M.; Kamalam, B.S.; Sharma,

A.; Dash, P. & Akhtar, M.S. 2021. Intra-annual changes in reproductive indices of male and female Himalayan snow trout, *Schizothorax richardsonii* (Gray, 1832). Aquaculture Research 52(1): 130-141.

- Dar, S.A.; Kaur, H. & Chishti, M.Z. 2017. First record of myxozoan parasites from fresh water fishes of Jammu and Kashmir and their pathogenecity. Microbial Pathogenesis 105: 138-144.
- Dar, S.A.; Kaur, H. & Chishti, M.Z. 2017. *Myxobolus chushi* n. sp. (Myxozoa: Myxosporea) parasitizing *Schizothorax niger* (Heckel), a native cyprinid fish from Wullar Lake in Kashmir Himalayas. Parasitology international 66(3): 272-278.
- Dar, S.A.; Kaur, H.; Chishti, M.Z. & Ahmad, F. 2016. Protozoan parasites in a coldwater cyprinid fish *Schizothorax niger* from Kashmir himalayas. Bulletin of Pure & Applied Sciences-Zoology 35: 65-71.
- Das, D. & Nag, T.C. 2008. Morphology of adhesive organ of the snow trout *Schizothorax richardsonii* (Gray, 1832). Italian Journal of Zoology 75(4): 361-370.
- Das, D.N.; Abujam, S. & Singh, A.D. 2019. Research Trends on Fish & Fisheries in Mountain Waters of Eastern Himalayan Region.
- Das, S.M. & Koul, B.L. 1965. The gross anatomy of gonads and fecundity of four fishes of Kashmir. Kashmir Science 2(1-2): 64-76.
- DeSalle, R. & Goldstein, P. 2019. Review and interpretation of trends in DNA barcoding. Frontiers in Ecology and Evolution 7: 302.
- De Silva, S.S.; Abery, N.W. & Nguyen, T.T. 2007. Endemic freshwater finfish of Asia: distribution and conservation status. Diversity and Distributions 13(2): 172-184.
- Douglas, M.E. & Matthews, W.J. 1992. Does morphology predict ecology? Hypothesis testing within a freshwater stream fish assemblage. Oikos 213-224.
- Dudgeon, D.; Arthington, A.H.; Gessner, M.O.; Kawabata, Z.I.; Knowler, D.J.; Lévêque, C.; Naiman, R.J.; Prieur-Richard, A.H.; Soto, D.; Stiassny, M.L. & Sullivan, C.A. 2006. Freshwater biodiversity: importance, threats, status and conservation challenges. Biology 81(2): 163-82.
- Everard, M. & Kataria, G. 2011. Recreational angling markets to advance the conservation of a reach of the Western Ramganga River, India. Aquatic Conservation 21(1): 101-8.
- Fischer, J.; Lindenmayer, D.B. & Fazey, I. 2004. Appreciating ecological complexity: habitat contours as a conceptual landscape model. Conservation Biology 18(5): 1245-1253.
- Fricke, R. (2023). Eschmeyer's Catalog of Fishes: Genera, Species, References 2023. California Academy of

Sciences. USA. En línea: https://bit. ly/3FNIWbe, 22(02).

- Gandotra, R.; Shanker, R. & Singh, D. 2009. Studies on fecundity of snow trout *Schizothorax richardsonii* (Gray) from the lotic bodies of Rajouri district (J&K). Current World Environment 4(1): 127.
- Goel, C.; Sati, J.; Barat, A.; Patiyal, R.S.; Ali, S. & Sahoo, P.K. 2014. Complete mitochondrial genome organization of *Schizothorax richardsonii* (Gray, 1832). Mitochondrial DNA 25(3): 171-172.
- Goswami, M.; Sharma, B.S.; Bahuguna, S.N.; Nagpure, N.S. & Lakra, W.S. 2013. A SRCF cell line from snowtrout, *Schizothorax richardsonii*: development and characterization. Tissue and Cell 45(3): 219-226.
- Gupta, N.; Rajvanshi, A. & Badola, R. 2017. Climate change and human–wildlife conflicts in the Indian Himalayan biodiversity hotspot. Current Science 113(5): 846-7.
- Gupta, N.; Sivakumar, K.; Mathur, V.B. & Chadwick, M.A. 2015. Terrestrial protected areas and managed reaches conserve threatened freshwater fish in Uttarakhand, India. Parks 21(1): 89-101.
- Hagedorn, M.M.; Daly, J.P.; Carter, V.L.; Cole, K.S.; Jaafar, Z.; Lager, C.V. & Parenti, L.R. 2018. Cryopreservation of fish spermatogonial cells: the future of natural history collections. Scientific reports 8(1): 1-11.
- Hassan, M.A.; Ferrer-Boix, C.; Cienciala, P. & Chartrand, S. 2017. Sediment transport and channel morphology implications for fish habitat. Open Channel Hydraulics, River Hydraulic Structures and Fluvial Geomorphology pp. 322-348.
- Hayden, B.; Harrod, C. & Kahilainen, K.K. 2014. Lake morphometry and resource polymorphism determine niche segregation between cool-and cold-wateradapted fish. Ecology 95(2): 538-552.
- He, D.; Chen, Y.; Chen, Y. & Chen, Z. 2004. Molecular phylogeny of the specialized schizothoracine fishes (Teleostei: Cyprinidae), with their implications for the uplift of the Qinghai-Tibetan Plateau. Chinese Science Bulletin 49(1): 39-48.
- Heino, J.; Erkinaro, J.; Huusko, A. & Luoto, M. 2016. Climate change effects on freshwater fishes, conservation and management. Cambridge University Press: Cambridge, pp. 76-106
- Indu, S. 2010. Bionomical notes on ecological, feeding and breeding aspects of *Schizothorax richardsonii* (Gray), Himachal Pradesh, India. Annals of Forestry 18(2): 311-316.
- IUCN. 2020. The IUCN Red List of Threatened Species. Version 2020-1.
- Jackson, D.C. & Marmulla, G. 2001. The influence of

dams on river fisheries. FAO fisheries technical paper 419: 1-44

- Jan, M.; Jan, U. & Shah, G.M. 2014. Studies on fecundity and Gonadosomatic index of *Schizothorax plagiostomus* (Cypriniformes: Cyprinidae). Journal of Threatened Taxa 6(1): 5375-5379.
- Jan, N.A. & Das, S.M. 1970. Qualitative and quantitative studies on the food of eight fishes of Kashmir valley. Ichthyologica 10(1-2): 20-26.
- Jen, L. 2008. Systematics and biology of snow trout *schizothorax richardsonii* gray 1832 from some water bodies of Nagaland.
- Jhingran, V.G. 1991. Fish and fisheries of India. Hindustan Publishing Corp, New Delhi 727.
- Jhingran, V.G. & Sehgal, K.L. 1978. Coldwater fisheries of India. Coldwater fisheries of India.
- Johal, M.S.; Sharma, A.; Dubey, V.K.; Johnson, J.A.; Rawal, Y.K. & Sivakumar, K. 2021. Invasive brown trout Salmo trutta induce differential growth strategies in the native snow trout *Schizothorax richardsonii* of Himalaya: Are natives in unaltered rivers better at picking the gauntlet of invasion?. Journal of Applied Ichthyology 37(5): 723-734.
- Joshi, K.D. 2001. Brood stock rearing of threatened *Schizothorax richardsonii* in farm conditions. "Workshop and captive breeding of prioritised cultivable and ornamental fishes for commercial utilisation and conservation". National bureau of fish genetics resources, Lucknow 22: 12.
- Joshi, K.D. (2011). Artificial breeding and rearing of Schizothorax richardsonii (Gray). Indian Journal of Fisheries 51(2): 233-237.
- Joshi, K.D.; Alam, M.A.; Jha, D.N.; Srivastava, K.; Srivastava, S.K.; Kumar, V. & Sharma, A.P. 2017. Studies on ecology, fish diversity and fisheries of Ken–Betwa Rivers (India): Proposed for interlinking. Aquatic Ecosystem Health & Management 20(1-2): 71-85.
- Joshi, K.D.; Das, S.C.S.; Khan, A.U.; Pathak, R.K. & Sarkar, U.K. 2016. Reproductive biology of snow trout, *Schizothorax richardsonii* (1832) in a tributary of River Alaknanda, India and their conservation implications. International Journal of Zoological Investigations 2: 109-114.
- Joshi, K.D.; Sunder, S.; Joshi, C.B. & Vass, K.K. 2005. Domestication and growth of *Schizothorax richardsonii* (Gray). Journal of the Inland Fisheries Society of India 37: 64-67.
- Jyoti, M.K. & Malhotra, Y.R. 1975. Seasonal variations in feeding of *Noenracheiluskmhmirensis* (Hora). Matsya 1: 53-58.
- Kamalam, B.S.; Mahija, J.; Baral, P.; Pandey, A.; Akhtar,

M. S.; Ciji, A. & Rajesh, M. 2019. Temperature and oxygen related ecophysiological traits of snow trout (*Schizothorax richardsonii*) are sensitive to seasonal changes in a Himalayan stream environment. Journal of thermal biology 83: 22-29.

- Kapila, R. & Mishra, D.P. 2006. Randomly amplified polymorphic DNA (RAPD) fingerprinting of *Schizothorax richardsonii* (Gray). Indian Journal of Fisheries 53(2): 219-224.
- Kapila, R.; Kapila, S. & Basade, Y. 2002. Impact of temperature variation on haematology and serum enzymes of *Schizothorax richardsonii* (Gray). Indian Journal of Fish 49(2): 187-192.
- Khanal, G.P., Tshering, S. & Dorji, T. 2023. Lengthweight relationship and relative condition factor of a migratory and a resident hill stream fish from hydropower-impacted tributaries of punatsangchhu river, Bhutan. Bhutan Journal of Animal Science 7(1), 90-99.
- Koundal, S. Dhanze, R.; Koundal, A. & Sharma, I. 2013. Relative gut length and gastro-somatic index of six hill stream fishes, Himachal Pradesh, India. Journal of Environment and Biosciences 27(1): 11-18.
- Koundal, S.; Koundal, A.; Sharma, I. & Dhanze, R. 2016. Mouth morphometry and body lengths with respect to the feeding habits of Hill stream fishes from Western Himalaya HP (India). International Journal of Fisheries and Aquatic Studies 4(3): 346-356.
- Lakra, W.S.; Mohindra, V. & Lal, K.K. 2007. Fish genetics and conservation research in India: status and perspectives. Fish Physiology and Biochemistry 33(4): 475-487.
- Li, P.; Xi, B.W.; Zhao, X. & Xie, J. 2017. *Myxobolus linzhiensis* n. sp. (Myxozoa: Myxobolidae) from the gill filament of *Schizothorax oconnori* Lloyd (Cyprinidae: Schizothoracinae) in Tibet, China: morphological and molecular characterization. Parasitology research 116(11): 3097-3103.
- Linke, S.; Kennard, M.J.; Hermoso, V.; Olden, J.D.; Stein, J. & Pusey, B. J. 2012. Merging connectivity rules and large-scale condition assessment improves conservation adequacy in river systems. Journal of Applied Ecology 49(5):1036-1045.
- Lohani, V.; Pant, B.J.; Pandey, N.N. & Ram, R.N. 2020. Morphological account of *Schizothorax richardsonii* population of river kosi and river Alaknanda. Pharma Innovation 9(3): 30-33
- Ma, Q.; He, K.; Wang, X.; Jiang, J.; Zhang, X. & Song, Z. 2020. Better resolution for Cytochrome b than Cytochrome c Oxidase subunit I to identify Schizothorax species (Teleostei: Cyprinidae) from the

Tibetan Plateau and its adjacent area. DNA and cell biology 39(4): 579-598.

- Madhwal, B.P.; Singh, H.R. & Chopra, A.K. 1984. Monthly variation in the dietary habit of *schizothorax-richardsonii* (gray) from the river yamuna in garhwal Himalayas. Indian journal of animal sciences 54(11): 1108-1110.
- Magurran, A.E. 2009. Threats to freshwater fish. Science 325(5945): 1215-1216.
- Majhi, S.K.; Das, S.K. & Rajkhowa, D. 2013. Effects of elevated water temperature on tolerance and stress in Chocolate mahseer *Neolissochilus hexagonolepis*: implications for habitat restoration and conservation. Current Science 379-383.
- Mali, J.D. & Chutia, P. 2017. Impact of Ranganadi dam on Socio-economic condition of fisher community in the downstream of Ranganadi river dam, NE India. International Journal of Engineering Science Invention 6(10): 19-23.
- Malik, D.S.; Sharma, A.K. & Vishal, K. (2021). Lengthweight relationships of coldwater fish species from the different stretches of upper Ganga basin in Garhwal region of Uttarakhand. Annals of Agri Bio Research 26(1), 98-100.
- Mallik, S.K.; Shahi, N.; Das, P.; Pandey, N.N.; Haldar, R.S.; Kumar, B.S. & Chandra, S. 2015. Occurrence of *Ichthyophthirius multifiliis* (White spot) infection in snow trout, *Schizothorax richardsonii* (Gray) and its treatment trial in control condition. Indian Journal of Animal Research 49(2): 227-230.
- Mallik, S.K.; Shahi, N.; Pandey, N.N.; Haldar, R.S. & Pande, A. 2010. Occurrence of fish louse (Argulus sp.) on Indian snow trout (*Schizothorax richardsonii*) and golden mahseer (*Tor putitora*) in subtropical Himalayan Lake of Bhimtal, Uttarakhand, India. The Indian Journal of Animal Sciences 80: 1152-1156.
- Margules, C.R. & Pressey, R.L. 2000. Systematic planning for biodiversity conservation. Nature 405: 243-253.
- Mehmood, S.; Ahmed, I. & Ali, M.N. (2021). Lengthweight relationship, morphometric and meristic controlling elements of three freshwater fish species inhabiting North Western Himalaya. Egyptian Journal of Aquatic Biology & Fisheries 25(6): 243-257.
- Meyer, A. 1987. Phenotypic plasticity and heterochrony in *Cichlasoma managuense* (Pisces, Cichlidae) and their implications for speciation in cichlid fishes. Evolution 41(6): 1357-1369.
- Mijkherjee, M.; Praharaj, A. & Das, S. 2002. Conservation of endangered fish stocks through artificial propagation and larval rearing technique in West Bengal, India. Aquaculture Asia 7(2): 8-11.

- Mir, J.I.; Shabir, R. & Mir, F.A. 2012. Length-weight relationship and condition factor of *Schizopyge curvifrons* (Heckel, 1838) from River Jhelum, Kashmir, India. World Journal of Fish and Marine Sciences 4(3): 325-329.
- Mir, S. 1979. Studies on the biology of Oreinus plagiostomus (Mc clleland) (Doctoral dissertation, Ph. D. thesis, Kashmir University, Kashmir).
- Mishra, K.S. 2003. An aid to commercial fisheries. Narendra pub house India
- Misra, M. 1982. Studies on fishery biology of *Schizothorax richardsonii* (Gray)-an economically important food fish of Garhwal Himalaya (Doctoral dissertation, Ph. D. thesis submitted to Garhwal University, Srinagar, Garhwal).
- Mohan, M. 2005. Spawning biology of snow trout, *Schizothorax richardsonii* (gray) from River Gaula (Kumaon, Himalayas). Indian Journal of Fisheries 52(4): 451-457.
- Mohan, M.; Bhanja, S. & Basade, Y. 2009. Performance of chitin incorporated diet on the indigenous Kumaon Himalayan fishes: snow trout, *Schizothorax richardsonii* (Gray) and golden mahseer, *Tor putitora* (Hamilton). Indian Journal of Fisheries 56(2): 135-137.
- Moyle, P.B. 1995. Conservation of native freshwater fishes in the Mediterranean-type climate of California, USA: a review. Biological Conservation 72(2): 271-279.
- Nag, T.C. & Bhattacharjee, J. 2002. Retinal cytoarchitecture in some mountain-stream teleosts of India. Environmental Biology of Fishes 63(4): 435-449.
- Negi, R.K. & Rajput, V. 2012. Fish diversity in two lakes of kumaon Himalaya Uttarakhand, India. Research Journal of Biology 2(5): 157-161.
- Negi, R.K., & Negi, T. 2010. Analysis of morphometric characters of *Schizothorax richardsonii* (Gray 1832) from the Uttarkashi District of Uttarakhand State, India. Journal of Biological Sciences 10(6): 536-540.
- Negi, R.K.; Tyagi, P. & Joshi, B.D. 2015. Morphometric Analysis of *Schistura montanus* from the Garhwal and Kumaun regions in Uttarakhand State, India. World Journal Fish and Marine Sciences 7(5): 400-403.
- Neupane, S. (2019). Length-Weight Relationship, Condition Factor and Stomach Content Analysis of Snow Trout from Khanikhola, Kavrepalanchok (Doctoral dissertation, Central Department of Zoology).
- Oberdoff, T.; Guégan, J.F. & Hugueny, B. 1995. Global scale patterns of fish species richness in rivers. Ecography 18(4): 345-352.
- Olden, J.D.; Kennard, M.J.; Leprieur, F.; Tedesco, P.A.;

Winemiller, K. O. & García-Berthou, E. 2010. Conservation biogeography of freshwater fishes: recent progress and future challenges. Diversity and Distributions 16(3): 496-513.

- Pandey, S.K. & Nautiyal, P. 1997. Statistical evaluation of some meristic and morphometric characters of taxonomic significance in *Schizothorax richardsonii* (Gray) and *Schizothorax plagiostomus* (Heckel). Indian Journal of Fisheries 44(1): 75-79.
- Pandey, S.K.; Nautiyal, P. (1997). Statistical evaluation of some meristic and morphometric characters of taxonomic significance in *Schizothorax richardsonii*. Indian Journal of Fisheries 44 (1): 75-79. CiteSeerX 10.1.1.972.5967.
- Pandit, M.K. & Grumbine, R.E. 2012. Potential effects of ongoing and proposed hydropower development on terrestrial biological diversity in the Indian Himalaya. Conservation Biology 26(6): 1061-1071.
- Petr, T. & Swar, D.B. 2002. Cold water fisheries in the trans-Himalayan countries. Food & Agriculture Organisation.
- Pikitch, E.K.; Santora, C.; Babcock, E.A.; Bakun, A.; Bonfil, R.; Conover, D.O.; Dayton, P.; Doukakis, P.; Fluharty, D.; Heneman, B.; Houde, E.D.; Link, J.; Livingston, P.A.; Mangel, M.; Mcallister, M.K.; Pope, J. & Sainsbury, K. J. 2004. Ecosystem-based fishery management. Science 305(5682): 346-347.
- Qadri, M.Y.; Mir, S. & Yousuf, A.R. 1983. Breeding biology of *Schizothorax richardsonii* gray and hard. Journal of the Indian Institute of Science 64(6): 73.
- Rai, A.K.; Pradhan, B.R.; Basnet, S.R. & Sawr, D.B. 2002. Present status of snow trout in Nepal. FAO Fisheries Technical Paper 213-220.
- Raina, H.S. & Petr, T. 1999. Coldwater fish and fisheries in the Indian Himalayas: lakes and reservoirs. Fish and fisheries at higher altitudes: Asia 64-88.
- Raizada, M.N.; Jain, K.K. & Raizada, K.K. 1983. Monthly variations in the hematocrit values (PCV) in a teleost, *Cirrhinus mrigala* (Ham.). Journal of Comparative Physiology 8: 196-198.
- Raj Kumari, Sharma, B.K., Sharma, L.L & Upadhyay, B.
 2006. Length-weight relationship and condition factor of Catla catla (Ham.) and Labeo rohita (Ham.) from Daya Reservoir Udaipur (Rajasthan). 1. Inland Fisheries Society of India 38 (1): 72-76.
- Rajesh, M.; Kamalam, B.S.; Ciji, A.; Akhtar, M.S.;
 Pandey, N.; Gupta, S.; Sarma, D.; Sahu, N.P. & Singh,
 A.K. 2019. Molecular characterisation and
 transcriptional regulation of muscle growth regulatory
 factors myogenin and myogenic factor 6 in the TransHimalayan cyprinid fish Schizothorax

richardsonii. Comparative biochemistry and physiology Part A, Molecular & Integrative Physiology 231: 188-200.

- Rajput, V.; Johnson, J.A. & Sivakumar, K. 2013.
 Environmental effects on the morphology of the Snowtrout *Schizothorax richardsonii* (Gray, 1832). TAPROBANICA: The Journal of Asian Biodiversity 5: 102-110.
- Rajvanshi, A.; Arora, R.; Mathur, V.B.; Sivakumar, K.; Sathyakumar, S.; Rawat, G.S.; Johnson, J.A.; Ramesh, K.; Dimri, N. & Maletha, A. 2012. Assessment of cumulative impacts of hydroelectric projects on aquatic and terrestrial biodiversity in Alaknanda and Bhagirathi basins, Uttarakhand. Wildlife Institute of India, Technical Report 203.
- Rampal, C. 1967. Studies on the life history of five Kashmir fishes together with fecundity and breeding habits. PhD thesis, University of Kashmir, Kashmir
- Rathore, S.S.; Chandravanshi, A.; Chandravanshi, P. & Jaiswal, K. 2016. Review on cryopreservation in fisheries science: Applications and perspectives. International Journal of Science and Research 5: 573-579.
- Raut, S.; Gupta, N.; Nautiyal, P. & Everard, M. 2020. Reestablishment of fish passage for conserving threatened migratory species of West-Indian Himalayas. River Research and Applications 36: 314-317.
- Regmi, B. 2019. Phylogenomics and geometric morphometrics defne species focks of snow trout (Teleostei: *Schizothorax*) in the Central Himalayas. PhD dissertation, University of Arkansas 139.
- Richter, B.D.; Braun, D.P.; Mendelson, M.A. & Master, L.L. 1997. Threats to imperiled freshwater fauna. Conservation Biology 11: 1081-1093. https://doi.org/10.1046/j.1523-1739.1997.96236.x
- Rodríguez, M.A. 2002. Restricted movement in stream fish: the paradigm is incomplete, not lost. Ecology 83: 1-13. https://doi.org/10.1890/0012-9658(2002)083
- Roy, N.K.; Raymajhi, A.; Pradhan, N. & Wagle, S.K. 2011. Reproductive performance of domesticated asala (*Schizothorax richardsonii*) in captive environment at godawari, Lalitpur. (eds.):100
- Sarkar, U.K. & Bain, M.B. 2000. Priority habitats for the conservation of large river fish in the Ganga River basin. Aquatic Conservation: Marine and Freshwater Ecosystems 17: 349-359.
- Sarkar, U.K.; Pathak, A.K. & Lakra, W.S. 2008. Conservation of freshwater fish resources of India: new approaches, assessment and challenges. Biodiversity and Conservation 17: 2495-2511.
- Sarkar, U.K.; Pathak, A.K.; Sinha, R.K.; Sivakumar, K.; Pandian, A.K. & Lakra, W.S. 2012. Freshwater fish

biodiversity in the river Ganga (India): changing pattern, threats and conservation perspectives. Reviews in Fish Biology and Fisheries 22: 251-272.

- Sarma, D.; Akhtar, M.S. & Singh, A.K. 2018. Coldwater fisheries research and development in India. Aquaculture in India 93-133.
- Sehgal, K.L. & Sar, C.K. 1989. Impact of construction and completion of Beas-Sutlej (BSL) Project on coldwater fisheries of R. Beas in Himachal Pradesh. In: National Workshop on Research and Development Needs of Coldwater Fisheries in India, Haldwani. Abstract No.14
- Sehgal, K.L. 1988. Ecology and Fisheries of Mountain Streams of North-Western Himalayas. Doctoral dissertation, Thesis for the award of DSc degree, University of Meerut, India.
- Sehgal, S.L. 1990. Ecology, abundance and distribution of fish life in the northwestern Himalayan streams. Recent Trends in Limnology 287-299.
- Shabanum, Sharma, I. & Dhanze, R. 2016. The significance of osteology in identification of *Schizothorax* species. International Journal of Fisheries and Aquatic Studies 4: 442-447.
- Sharma, A.; Dubey, V.K.; Johnson, J.A.; Rawal, Y.K. & Sivakumar, K. 2021a. Is there always space at the top? Ensemble modeling reveals climate-driven highaltitude squeeze for the vulnerable snow trout *Schizothorax richardsonii* in Himalaya. Ecological Indicators 120: 106900.
- Sharma, A.; Dubey, V.K.; Johnson, J.A.; Rawal, Y.K. & Sivakumar, K. 2021b. Introduced, invaded and forgotten: allopatric and sympatric native snow trout life-histories indicate brown trout invasion effects in the Himalayan hinterlands. Biological Invasions 5: 1497-515.
- Sharma, I. & Mehta, H.S. (2010). Studies on snow trout Schizothorax richardsonii (Gray) in river Beas and its tributaries (Himachal Pradesh), India. Zoological Survey of India 323: 1-69.
- Sharma, I. 2018. Status of trout fishes versus climate change in Himachal Pradesh, North Western Himalaya. International Journal of Fisheries and Aquatic Studies 6: 424-426.
- Sharma, I. 2019. Ichthyodiversity of Beas River system north western Himalaya (HP), India. Journal of Environment and Bio- sciences 33(1): 11-17.
- Sharma, N.K.; Akhtar, M.S.; Pandey, N.; Singh, R. & Singh, A.K. 2015. Seasonal variation in thermal tolerance, oxygen consumption, antioxidative enzymes and non-specific immune indices of Indian hill trout, *Barilius bendelisis* (Hamilton, 1807) from central Himalaya, India. Journal of Thermal Biology 52: 166-

176.

- Shekhar, C. 1990. Biology of *Oreinus richardsonii* (Grey & Hard) from Neeru-Nallah in Bhaderwah with a view to formulation of species conservation plan. PhD Thesis, University of Jammu, India.
- Shekhar, C.; Malhotra, Y.R. & Dutta, S.P.S. 1993. Food and feeding habits of *Schizothorax richardsonii* (Gray and Hard) inhabiting Neeru nullah, Bhaderwah, Jammu. Journal of the Indian Institute of Science 73: 247-251.
- Shen, Y.; Hubert, N.; Huang, Y.; Wang, X.; Gan, X.; Peng, Z.; & He, S. 2019. DNA barcoding the ichthyofauna of the Yangtze River: Insights from the molecular inventory of a mega-diverse temperate fauna. Molecular Ecology Resources 19(5): 1278-1291.
- Shrestah, T.K. 2002. Cold water fisheries development in Nepal. FAO Fisheries Technical Paper 47-58.
- Singh, A.K. 2016. Fish Diversity of Himalayan Region, India for Sustainable Development. Uttar Pradesh State Biodiversity Board 140-144.
- Singh, A.K. 2019. Coldwater Fisheries in India: Priorities, Policy, Institutional Support and Challenges. Journal of Advanced Agricultural Technologies 3(2): 152-6.
- Singh, A.K.; Kumar, P. & Ali, S. 2014. Ichthyofaunal diversity of the Ganges River system in central Himalavas. India: conservation status and priorities. Rivers for Life-Proceedings of the International Symposium on River Biodiversity: Ganges-Brahmaputra-Meghna River System, Ecosystems for Life, A Bangladesh-India Initiative, IUCN, International Union for Conservation of Nature 208-214.
- Singh, H. & Bisht, I. 2017. Comparative ultrastructural study of general body epidermis of the hill-stream fishes; *Botia almorhae* (Teleosti: Botiidae), *Homaloptera brucei* (Teleostei: Balitoridae) and *Schizothorax richardsonii* (Teleostei: Cyprinidae). International Journal of Aquatic Biology 5(6): 370-374.
- Singh, H. 2018. Habitat fragmentation due to Tehri dam construction reduces fish species diversity among Bhilangana River, Uttrakhand, India. Asian Journal of Animal Sciences 13(2): 62-7.
- Singh, S. 1990. The lotic water fisheries of Kumaun Himalaya. Himalaya Environment Resources and Development 276-284.
- Sivakumar, K. 2008. Species richness, distribution pattern and habitat use of fishes in the Trans-Himalayas, India. Electronic Journal of Ichthyology 1: 31-42.
- Sivaraman, G.K.; Barat, A.; Ali, S.; Joshi, S. & Mahanta, P.C. 2010. Phylogenetic Analysis of Coldwater Fish

Species (Cyprinids) of India using targeted mtDNA and RAPD-PCR markers. The IUP Journal of Genetics & Evolution 3: 42-49.

- Sunder, S. 1985. Food and feeding habits of *Puntius conchonius* (Ham) from Dal Lake, Kashmir. Geobios 4: 131-136.
- Swar, D.B. 1990. Effect of impoundment on the indigenous fish population in Indrasarobar reservoir, Nepal (No. IDRC--291E)
- Talwar, C.; Nagar, S.; Lal, R. & Negi, R.K. 2018. Fish gut microbiome: current approaches and future perspectives. Indian Journal of Microbiology Research 58: 397-414.
- Talwar, C.; Singh, A.K.; Choksket, S.; Korpole, S.; Lal, R. & Negi, R.K. 2020. *Salinicoccuscyprini* sp. nov., isolated from the gut of mirror carp, *Cyprinuscarpio* var. *specularis*. International Jjournal of Systematic and Evolutionary Microbiology 70: 4111-4118.
- Talwar, P.K. & Jhingran, G.A. 1991. Inland fisheries of India and adjacent countries, vol. I and II. New Delhi: Oxford and IBH Publishing Company Pvt. Limited, New Delhi, 1158
- Tedesco, P.A.; Beauchard, O.; Bigorne, R.; Blanchet, S.; Buisson, L.; Conti, L.; Cornu, J.F.; Dias, M.S.; Grenouillet, G.; Hugueny, B. & Jézéquel, C. 2017. A global database on freshwater fish species occurrence in drainage basins. Scientific data 4: 1-6.
- Thapliyal, M.; Bahuguna, S.N. & Thapliyal, A. 2019. Adaptive skill of *Schizothorax* sp. of river Alaknanda under long term pressure of urbanization and anthropogenic activities in Garhwal Himalaya. Environment conservation journal 20: 157-163.
- Thapliyal, M.; Bahuguna, S.N.; Chandra, T. & Thapliyal, A. 2012. An account of ova maturation and Gonado-Somatic Index (GSI) of snow trout, *Schizothorax richardsonii* (Gray) of Garhwal Himalaya. Journal of Environment and Bio- sciences 26: 43-50.
- Uzma, S.; Bisht, H.C.S. & Pandey, N.N. 2020. Effects of experimental infection with *Aeromonas hydrophila* on different blood parameters and hematopoietic tissue in *Schizothorax richardsonii*. Journal of Experimental Zoology 23: 173-178.
- Vishwanath, W. 2010. *Schizothorax richardsonii* (errata version published in 2018) The IUCN Red List of Threatened Species 2010: e.T166525A135873256.
- Wadia, D.N. 1931. The syntaxis of the northwest Himalaya: its rocks, tectonics and orogeny. Record of Geological Survey of India 65(2): 189-220.
- Wagle, S.K. 2014. Studies on gonadosomatic index, fecundity and hatchability of domesticated stock of asala *Schizothorax richardsonii* (Gray) from Nallu

River of Lalitpur District. Our Nature 12: 19-27.

- Wagle, S.K.; Shrestha, M.K.; Rai, S. & Pandit, N.P. 2016. Effect of different protein levels on feed utilization, growth and body composition on fry of snow trout *Schizothorax richardsonii*. Yeast 1:3.
- Ward, R.D.; Zemlak, T.S.; Innes, B.H.; Last, P.R. & Hebert, P.D.N. 2005. DNA Barcoding Australia's fish species. Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences 360(1462): 1847-1857.
- Zhang, C.; Tong, C.; Ludwig, A.; Tang, Y.; Liu, S.; Zhang, R.; Feng, C.; Li, G.; Peng, Z. & Zhao, K. 2018. Adaptive evolution of the Eda gene and scales loss in schizothoracine fishes in response to uplift of the Tibetan plateau. International Journal of Molecular Science 19(10): 2953.
- Zhang, J.; Chen, Z.; Zhou, C.; & Kong, X. 2016. Molecular phylogeny of the subfamily Schizothoracinae (Teleostei: Cypriniformes: Cyprinidae) inferred from complete mitochondrial genomes. Biochemical Systematics and Ecology 64: 6-13.
- Zhou, S.; Smith, A.D.; Punt, A.E.; Richardson, A.J.; Gibbs, M.; Fulton, E.A.; Pascoe, S.; Bulman, C.; Bayliss, P. & Sainsbury, K. 2010. Ecosystem-based fisheries management requires a change to the selective fishing philosophy. Proceedings of the National Academy of Sciences of the United States of America 107: 9485-9489.

مقاله مروري

آرایهشناسی، پراکنش، زیستشناسی و حفاظت گونه آسیب پذیر قزل آلای برفی، (پر توبالگان: کپورماهیان: خواجو ماهیان)در مناطق هیمالیا و زیرحوضه آن: مروری

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چکیده: ماهیها جزء مهمی از محیط آبی هستند و در حمایت و حفظ کل سیستم اکولوژیک حائز اهمیت هستند، در نتیجه سیستمهای آب شیرین را انعطاف پذیرتر و پایدارتر می کنند. ساختار جامعه ماهی به فعل و انفعالات زیستی و متغیرهای غیر زنده بستگی دارد. ساختار جوامع ماهیان وابسته به فعل و انفعالات زیستی و متغیرهای غیر زیستی است. طی ۵ تا ۱۰ سال گذشته، حامیان محیط زیست و ماهیگیران تجاری، به دلیل ورود گونههای غیربومی، سدسازی، فعالیتهای انسانی، ماهیگیری غیرقانونی و صید بی رویه، کاهش شدیدی را در شیلات آب شیرین نشان دادهاند. Schizothorax richardsonil یک ماهی آب شیرین بسیار ارزشمند است که در دامنههای هیمالیا حیات دارد. کاهش شدیدی را در شیلات آب شیرین نشان دادهاند. Schizothorax richardsonil یک ماهی آب شیرین بسیار ارزشمند است عنوان شده است. با مقاومت در برابر متغییرهای زیستی و غیر زیستی، دامنه پراکنش این گونه در نقاط مختلف از لحاظ ارتفاع از سطح دریا زیاد شده است و با توجه به عنوان شده است. با مقاومت در برابر متغییرهای زیستی و غیر زیستی، دامنه پراکنش این گونه در نقاط مختلف از لحاظ ارتفاع از سطح دریا زیاد شده است و با توجه به عنوان شده است. با مقاومت در برابر متغییرهای زیستی و غیر زیستی، دامنه پراکنش این گونه در نقاط مختلف از لحاظ ارتفاع از سطح دریا زیاد شده است و با توجه به عنوان شده است. با مقاومت در برابر متغییرهای زیستی و غیر زیستی، دامنه پراکنش این گونه در نقاط مختلف از لحاظ ارتفاع از سطح دریا زیاد شده است و با توجه به عنوان شده است. با مقاومت در برابر متغییرهای زیستی و غلهراً دامنه توزیع فعلی آن در طول زمان از بین می رود. در مناطق هند-هیمالیا، اقدامات حفاظتی وجود ندارد، در نتیجه، علی رغم اهمیت اکولوژیک، ماهی قزلآلای برفی براساس لیست 2020 IUCN وضعیت آسیب پذیری پیدا کرده است. هدف از این بررسی، ارائه زیستشناسی توصیفی و بحث در مورد تأثیر فعالیتهای انسانی بر ساختار جمعیت می و مفاظت از آن است.